

Original Research Article

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## Clinico-Pathological and Compositional Changes in Milk of Mastitis Cows

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### ABSTRACT

The present study was aimed to have an in-depth study on haemato-biochemical alterations in mastitis cows along with compositional changes in mastitis milk. There was an overall incidence of 41.88 % of mastitis screened on basis of clinical signs, SCC and MCMT with 77.56 % subclinical mastitis and 22.54 % of clinical mastitis. Hematology of mastitis cows revealed anemia with significant decrease in Hb, TEC and PCV, leucocytosis with neutrophilia, eosinophilia and lymphopaenia. Significant increase in Total Protein, AST, Na<sup>+</sup> and K<sup>+</sup> concentrations and a significant decrease in Glucose, Ca<sup>+</sup> and Phosphorous concentrations was seen in affected cows. Significant (p<0.05) increase in pH, Whey Protein, Na<sup>+</sup> and electrical conductivity and a significant (p<0.05) decrease in specific Gravity, acidity, Fat, SNF and total protein value observed mastitis milk. Tetracycline was found as the most prevalent antibiotic residues. Histopathology of the representative tissue sample revealed the presence of ductal haemorrhages with adjacent fibrotic proliferation, polypoid fibrotic growth projecting into the lumen of teat canal, fibrosis along with inflammatory cells filled in ductal areas. Data generated during the study envisages understanding the disease severity for remedial therapy and assessing deterioration in the milk quality for food safety.

#### Keywords

Hematology,  
Histopathology, Mastitis,  
Milk composition, Serum  
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### Introduction

Mastitis is the inflammation of parenchyma of mammary gland characterized by physical, chemical, compositional and usually bacteriological changes in milk and

pathological changes in glandular tissues (Radostits *et al.*, 2000).

Alterations in hemato-biochemical parameters have been correlated to aid diagnosis as well as treatment. Early detection with suitable

tests, timely intervention with effective treatment and prevention is necessary to restrict the spread of mastitis (Guha and Gera, 2011). Patho-morphological changes in glandular tissue owing to mastitis results in compositional changes in milk leading to unsuitable for consumption.

Keeping in view of the high rate of prevalence as well as devastating production loss in relation with both quantity and quality of milk and possibility of permanent teat damage, this study is designed to have a detailed study on clinic-pathological changes associated in mastitis cows.

## **Materials and Methods**

### **Screening of cows to study incidence of mastitis**

The cows domesticated in and around Bhubaneswar were included in the present research programme randomly screened for detection of mastitis on the basis of history from the owner about reduced milk yield, presence / absence of abnormality in milk such as flakes, clots, and watery milk, bloods or gels or precipitates in milk, Presence / absence of clinical signs, somatic cell count of the milk samples (SCC) and Modified California Mastitis Test (MCMT).

During screening, the cows were grouped as sub-clinical and clinical mastitis basing on SCC and CMT score along with signs of inflammation in udder and gross abnormal composition of milk.

### **Collection of Milk Sample**

Milk samples were collected aseptically from the mastitis cows. Samples were labeled properly after collection and then transferred to the laboratory for further laboratory works like cytology and SCC besides CMT done instantly at the field at owners door step.

### **California Mastitis Test (CMT)**

The rapidity and amount of gel formation or precipitate is measured qualitatively to assess the severity. The test was done as per the standard procedures described by Schlam and Noorland (1957).

### **Somatic Cell Count (SCC)**

The procedure followed was according to the general principle of Prescott and Breed method as detailed by Schalm *et al.*, (1971). Cells from 25 fields were counted, averaged and multiplied by  $5 \times 10^5$  to get the total number of somatic cells/ml of milk.

### **Compositional changes in milk**

Alterations in milk composition in mastitis cows by comparing with the milk from normal but apparently healthy cows in relation to consistency, pH, specific gravity, fat%, SNF%, acidity, total protein, whey protein and  $\text{Na}^+$  concentrations were seen. The data obtained was analyzed by various statistical methods.

### **Antibiotic Residues in Mastitis Milk**

Ten mastitis milk samples selected for QuickSTAR strip reader for possibility regarding presence of any antibiotic residues.

### **Haemato-biochemical examinations**

Blood sample from the apparently healthy lactating cows as well as cows suffering from mastitis were subjected to detailed haemato-biochemical estimation. Serum samples were estimated in Automatic Biochemical Analyser (Turbo Chem 100) using commercially available ICHEM kits manufactured by Jeev diagnostics and the methods of analysis were as per the manufacturer's instruction. Hemoglobin percentage was estimated as per the Sahali's method.

### ***Histopathology of Mastitis Udder Tissue***

The representative tissue samples were collected by curetting from the blocked quarters of chronic mastitis cases (n=6) and processed for routine histopathology under routine H&E stain. Data generated in the study analyzed by using SAS software package.

### **Results and Discussion**

There was an overall prevalence of 41.88% out of 234 cows screened for mastitis on basis of clinical signs, SCC and MCMT. Average SCC of 2.5 – 4.0 lakhs/ml without any gross abnormality of udder considered as subclinical mastitis and more than 4.0 lakhs/ml of SCC with presence of clinical signs related to udder gross pathological changes considered as clinical mastitis. Below 2.5 lakhs/ml of SCC without any obvious changes in milk as well as udder considered as normal and apparently healthy but non-mastitis cows. Cows were screened for MCMT and scored accordingly 3+ i.e. (Distinct slime formation which adheres to the bottom of the paddle) of 17, 2+ (slime formation) i.e.19, 1+ (distinct precipitate but no tendency towards gel formation) i.e. 62 and 136, Negative (0) i.e. no evidence of a formation of a precipitate in the present study. Out of 98 cows screened positive for mastitis, 76 cows (77.56 %) were of subclinical mastitis and only 22 cows (22.54 %) were of clinical mastitis. Environmental factors such as climate, housing, and quality of indoor air, type of bedding / floor, stress and non-availability of nutrients were proven to be the predisposing factors for high incidence of the disease in this study as the cows housed in natural field conditions (Radostits *et al.*, 2007). The earlier researchers from Odisha reported a similar higher prevalence of subclinical mastitis i.e. of 43.57 % (Sahoo and Parida, 2002). Unawareness among the farmers about good managemental practices is considered for the

higher prevalence of subclinical mastitis in some localities as reported by Zeedan *et al.*, (2014).

### **Haematological examination**

Haematological parameters were evaluated in mastitis cows and compared with apparently healthy cows depicted in Table-1. There was a significant ( $p<0.05$ ) decrease in Hb, TEC and PCV with significant ( $p<0.05$ ) increase in TLC in mastitis cows in comparison to Control cows i.e. apparently healthy but mastitis negative cows. Differential leucocyte count (DLC) of the mastitis cows showed a significant ( $p<0.05$ ) increase in Neutrophil and Eosinophil. Sarvesha *et al.*, (2017) observed a significant increase in TLC and lymphopenia in mastitis infected animals. Significant reduction in Hb, TEC, and PCV values might lead to anaemia in affected animals. Increase in TLC along with neutrophilia and lymphopenia which may attribute to persistent infection in mastitis.

### **Serum biochemical examination**

The present study showed a significant ( $p<0.05$ ) increase in various serum biochemical parameters like Total Protein (TP), AST,  $\text{Na}^+$  and  $\text{K}^+$  concentrations and a significant decrease value for Glucose,  $\text{Ca}^+$  and Phosphorous concentrations in mastitis affected cows when compared with the apparently healthy but mastitis negative control cows as depicted in Table-2. Present finding corroborates with Babaei *et al.*, (2007); Kumar (2012) and Jain *et al.*, (2013). Zaki *et al.*, (2008) observed a non-significant decrease in the serum calcium and total protein values with no obvious changes in the serum phosphorus values in subclinical mastitis in comparison to the healthy control. The increased levels of various enzymes in the milk occur mainly due to the increased permeability of microcirculatory vessels in

inflamed areas along with the leakage from degenerated/necrotic parenchymal cells and leukocytes. Increase in Total Protein may be attributed to the decreased albumin levels after the immune response to the udder infection (Singh, 2000).

### **Compositional changes in milk**

Details of compositional changes of mastitis milk given in Table-3 showed a significant ( $p < 0.05$ ) increase in pH, Whey Protein,  $\text{Na}^+$  and electrical conductivity in mastitis milk in compared with normal but mastitis absent milk sample. There was a significant ( $p < 0.05$ ) decrease in Specific Gravity, acidity, Fat, SNF and total protein value in mastitis milk. This finding corroborates with Badran *et al.*, (1986) and Haggag *et al.*, (1991). Decrease in the specific gravity in mastitis affected cows attributed to the increase in chloride and decrease in the lactose contents. The study also reported that there is a positive correlation with the severity along with pH of the milk in mastitis. The decreasing trend in the specific gravity in mastitis milk owes to the reduced lactose contents and increased chlorides.

Higher levels of citrate and bicarbonate found during mastitis may be responsible for an elevated pH in mastitis milk as reported by Ogola *et al.*, (2007). The present findings regarding a decrease in Fat corroborates with the findings of Holdaway (1990) who stated that over a period of time, the total output of fat from a quarter is likely to be reduced, because of lower volume of milk. Also, the size of fat droplets is bigger relatively to that of intracellular gaps and so contained within the alveoli and consequently their concentration increases which corroborates with the findings of Auldish and Hubble, (1998). The study reported that the milk with high SCC is more susceptible to spontaneous lipolysis. Lipase enzymes produced by the

leukocytes at the mammary gland after infection are mainly responsible for the lipolysis of milk fat globule membranes, resulting in the breakdown of triglycerides and oxidation of fatty acids. Batavani *et al.*, (2007) concluded about elevated sodium, chloride, pH, albumin, lactate dehydrogenase (LDH) activity and immunoglobins in mastitis milk. In contrast, decreased values were found for calcium, inorganic phosphorus, potassium, alpha-lactalbumin and beta-lactoglobulin. Ogola *et al.*, (2007) reported significant increase in Na, Chloride and free fatty acids in mastitis milk samples while stated a significant lower value for lactose, casein-to-total protein, potassium and calcium. The results in these studies were linked to the reduced secretory activities of the mammary cells and increased permeability of the mammary epithelium. This can lead to the transfer of blood components to the milk. Hamadani *et al.*, (2013) reported a decrease in calcium concentrations in mastitis milk which is attributed to the disruption of casein. The ionic content of milk varies greatly in mastitis which plays a significant role in processing of milk samples and its quality. Potassium is found to be the most abundant mineral in milk, leaks out of milk through the paracellular pathway resulting a decrease in its concentration. Alternatively,  $\text{Na}^+$  found abundantly in blood, leaks into the milk resulting in an increasing concentration in mastitis milk. Due to heavy influx of blood constituents into the milk in infections i.e. in mastitis, the concentration of chloride is found to be elevated. The concentration of chloride in mastitis milk elevated probably due to the influx of blood constituent into the milk during infection conversely, sodium, found in the blood in high quantities, leaks into the milk increasing concentrations above normal (Auldish & Hubble, 1998). Increased electrical conductivity in mastitis milk in our study may be attributed to increase in  $\text{Na}^+$  concentrations which corroborates with Rajak *et al.*, (2015).

**Table.1** Mean  $\pm$  SE of Hematological alterations in cows affected with mastitis and apparently healthy cows reared in natural field conditions

Parameters	Control (n=10)	Mastitis (n=60)
Hb (gm%)	10.65 $\pm$ 0.27 <sup>a</sup>	8.8 $\pm$ 0.11 <sup>b</sup>
TLC	6960 $\pm$ 177.8 <sup>b</sup>	10670 $\pm$ 153.6 <sup>a</sup>
TEC	5.53 $\pm$ 0.12 <sup>a</sup>	4.66 $\pm$ 0.06 <sup>b</sup>
PCV	32.6 $\pm$ 0.84 <sup>a</sup>	27.3 $\pm$ 0.36 <sup>b</sup>
MCV	58.95 $\pm$ 0.28 <sup>a</sup>	58.07 $\pm$ 0.47 <sup>b</sup>
MCHC	32.68 $\pm$ 0.05 <sup>a</sup>	32.35 $\pm$ 0.08 <sup>b</sup>
Neutrophil	35.10 $\pm$ 1.47 <sup>b</sup>	50.96 $\pm$ 0.59 <sup>a</sup>
Lymphocyte	61.70 $\pm$ 1.52 <sup>a</sup>	44.31 $\pm$ 0.66 <sup>b</sup>
Eosinophil	1.70 $\pm$ 0.21 <sup>b</sup>	3.20 $\pm$ 0.24 <sup>a</sup>
Monocyte	1.50 $\pm$ 0.16	1.51 $\pm$ 0.06

NB: Mean bearing different superscript differs significantly at 5 % (P < 0.05)

**Table.2** Mean  $\pm$  SE of serum biochemical alterations in cows with mastitis and apparently healthy cows reared in natural field conditions

Parameters	Control (n=10)	Mastitis (n=20)
ALT	10.60 $\pm$ 0.01 <sup>a</sup>	10.17 $\pm$ 0.01 <sup>b</sup>
AST	88.70 $\pm$ 1.24 <sup>b</sup>	106.2 $\pm$ 1.60 <sup>a</sup>
Glucose	58.13 $\pm$ 0.16 <sup>a</sup>	56.43 $\pm$ 0.09 <sup>b</sup>
Total Protein	8.03 $\pm$ 0.07 <sup>b</sup>	9.54 $\pm$ 0.03 <sup>a</sup>
Ca	10.75 $\pm$ 0.15 <sup>a</sup>	8.11 $\pm$ 0.09 <sup>b</sup>
P	6.0 $\pm$ 0.13 <sup>a</sup>	5.01 $\pm$ 0.07 <sup>b</sup>
Na	139.9 $\pm$ 0.82 <sup>b</sup>	152.6 $\pm$ 1.05 <sup>a</sup>
K	3.98 $\pm$ 0.06 <sup>b</sup>	5.12 $\pm$ 0.07 <sup>a</sup>

NB: Mean bearing different superscript differs significantly at 5 % (P < 0.05)

**Table.3** Mean  $\pm$  SE of compositional changes in mastitis milk and normal milk

Parameters	Normal Milk (n=10)	Mastitis Milk (n=15)
pH	6.4 $\pm$ 0.03 <sup>b</sup>	6.9 $\pm$ 0.2 <sup>a</sup>
Specific gravity	1.030 $\pm$ 0.001 <sup>a</sup>	1.027 $\pm$ 0.003 <sup>b</sup>
Acidity	0.1316 $\pm$ 0.003 <sup>a</sup>	0.0853 $\pm$ 0.002 <sup>b</sup>
Fat	4.56 $\pm$ 0.08 <sup>a</sup>	3.31 $\pm$ 0.32 <sup>b</sup>
SNF	8.30 $\pm$ 0.05 <sup>a</sup>	7.6 $\pm$ 0.06 <sup>b</sup>
Total Protein	3.6 $\pm$ 0.008 <sup>a</sup>	3.5 $\pm$ 0.004 <sup>b</sup>
Whey Protein	0.77 $\pm$ 0.04 <sup>b</sup>	1.27 $\pm$ 0.03 <sup>a</sup>
Na	463.3 $\pm$ 21.22 <sup>b</sup>	868.9 $\pm$ 9.9 <sup>a</sup>
Electrical Conductivity	13.48 $\pm$ 0.05 <sup>b</sup>	14.54 $\pm$ 0.11 <sup>a</sup>

NB: Mean bearing different superscript differs significantly at 5 % (P < 0.05)

### **Antibiotic residues in milk**

Mastitis milk samples tested through QuickSTAR strip reader resulted about presence of tetracyclines in majority of milk sample as antibiotic residues. This might be due to low cost, easy availability and rampant use of this antibiotic in field conditions by majority of the vets and paravets.

### **Cytology of mastitis milk**

The cytology of mastitis milks showing presence of inflammatory cells along with numerous fat vacoules. There was clumping of cells as well as presence of lipid droplets in the stained smear. Ranjan *et al.*, (2008) reported similar findings in the cytology of mastitis milk. Presence of inflammatory cells along with fat vacuoles attributed to the active host's response to infection in the mammary gland.

### **Histopathology**

Histopathology of the representative tissue sample collected from chronic mastitis with blocked teat revealed the presence of ductal hemorrhages with adjacent fibrotic proliferation, polypoid fibrotic growth projecting into the lumen of teat canal, fibrosis along with ductal area filled with inflammatory cells.

There was presence of glandular lumen filled with erythrocytes. Some areas showed more proliferation of fibrotic cells along with congestion and hemorrhages and atrophic alveoli. In some cases, it was seen about extension of epidermal layer of the teat into fibrotic inner layers. Similar observations opined by several researchers such as Alawaa *et al.*, (2000); Green M and Bradley A (2004); Davidov *et al.*, (2011) and Hussain *et al.*, (2013). Fibrotic proliferation along with presence of RBCs and inflammatory cells in

glandular alveoli owes to animal's immune response against persistent infectious organisms.

Conclusively, present study of hemato-biochemical alterations in mastitis cows along with compositional changes in milk may be helpful to the clinician in assessing the severity of the affection as well as advising necessary therapy. Increased prevalence of subclinical mastitis without any obvious clinical signs with alterations in composition of milk pose a problem in dairy industry related to processing of milk products and its quality. Awareness among the farmers with routine screening of milk should be advocated to curb down the menace of this economic devastating disease in time to come.

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